










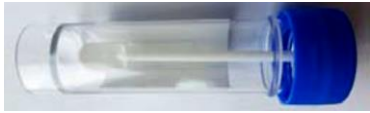


## BACTERIOLOGY SAMPLE CONTAINER GUIDE

All precious samples, such as CSF and respiratory samples including sputum, BALs etc **MUST NOT** be sent via the pneumatic tube. All other specimens, e.g., swabs, tips, blood cultures, faeces and urines can be sent via the pneumatic tube.

**\*\*All urgent samples need to be phoned to the laboratory prior to sending\*\***

Test	Container type	Comments
<b>MRSA</b> Nose & Groin/perineum only	 <p><b>Double eSwab</b> – pink cap, liquid media  <b>Manufacturer:</b> COPAN  <b>Lid:</b> Pink top  <b>Media:</b> Liquid Amies  <b>Media appearance:</b> Clear, colourless  <b>Swab:</b> 1 x white, 1 x pink</p>	Liquid eSwabs contain 1ml of liquid. No liquid should be discarded when collecting sample. <b>Samples with insufficient liquid will be rejected.</b>  <b>SEE: APPENDIX 1 for sample collection procedure</b>
<b>Wounds/throat swab/genital swabs</b> (Skin, superficial, not surgical), <b>Abscess or swab, deep-seated pus swab, post op wound swab, wound exudates</b> (all include MRSA)  <b>MRSA Screen – *for a rapid MRSA screening only result, please submit a separate swab</b>	 <p><b>Single eSwab</b> – pink cap, liquid media  <b>Manufacturer:</b> COPAN  <b>Lid:</b> Pink top  <b>Media:</b> Liquid Amies  <b>Media appearance:</b> Clear, colourless  <b>Swab:</b> 1 x white</p>	Collect the sample, insert the swab into the tube, snap off at the marked break point, discard the remaining shaft and recap the tube. <b>Do not discard any liquid.</b>
<b>Rapid/Routine Carbapenemase-Producing Enterobacteriaceae (CPE) Screen</b>	 <p><b>Double-head (duo) swab</b> – red cap, sponge with liquid media   <b>Manufacturer:</b> COPAN  <b>Lid:</b> Red top  <b>Media:</b> Foam sponge soaked in liquid media  <b>Media appearance:</b> Off-white sponge at base of container  <b>Swab:</b> 1 x 1 x double-head white</p>	Double headed red topped swab <b>Charcol swabs and wire samples are not suitable for this test and will be rejected.</b> Faecal material must be visible on the cotton tip of the swab otherwise the specimen will be rejected.
<b>Urethral swab</b>		

<p><b>Non- swab samples:</b>          Tissues, pus, Line Tips,          Aspirates, Sterile fluids,          Respiratory samples          (sputum, BALs, Pleural          fluid)</p>		<p>Universal container (30ml) – white          top, sterile container          Universal container (50ml) –          white/silver/yellow top, sterile          container  <b>Sterile fluids minimum volume:          1mL</b></p> <p>Tubes <b>MUST</b> be removed and          replaced with a securely sealed          screw cap</p>
<p><b>Sterile fluids for          culture:</b> CAPD/          peritoneal fluids          (Ascites), Joint Fluids          (Prosthetic &amp; Natural),          Stem Cell</p> <p><b>Pleural Fluids (Not          including pleural          drains)</b> are required to          be sent in a set of blood          culture bottles <b>AND</b> a          sterile universal</p>		<p>Sliver/blue top aerobic bottle</p> <p>Inoculate up to 10 mL to the bottle</p> <p>Adults: Sliver/blue top aerobic          bottle</p> <p>Paediatrics: Pink top Peds aerobic          bottle</p>
<p><b>Bronchial washings and          Bronchoalveolar          lavages</b></p>		<p>Tubes <b>MUST</b> be removed and          replaced with a securely sealed          screw cap</p>
<p><b>Urine</b>          Clean catch urine (CCU),          Mid stream urine          (MSU), Supra pubic          aspirate (SPA), Bladder          &amp; Catheter urine</p>	 <p>10 ml Sarsted urine Monovette tubes,          Minimal volume: 1mL</p>	<p>Urine samples must be collected in          a primary container and then          drawn into the 10ml Monovette          using the straw inside the          packaging.</p> <p><b>SEE: APPENDIX 2 for Urine          Monovette User Guide</b></p>

<b>Blood cultures</b>  Venous blood, arterial blood, peripheral blood, sterile fluids, stem cells  <b>Plastic bottles</b>		<b>BC Volume:</b> <b>Adults:</b> Inoculate between <b>8- 10 mL</b> to each bottle <b>Children:</b> Inoculate between <b>1- 3mL</b> <b>Neonates:</b> Inoculate 1-2 mL	Blood culture set is defined as one aerobic (sliver/blue top) and one anaerobic bottle (purple top)  For neonates and infants a single Peds aerobic bottle (pink top) is required.
<b>Faeces</b> (Routine culture, C. diff, H. pylori, Ova, Cysts and Parasites)			<b>Minimal volume</b> for liquid specimen: <b>2-3 mL</b>
<b>Perinasal swab</b> Bordetella pertussis (whooping cough)			Darcon with flexible wire shaft
<b>AFB including TB</b>  <b>Glass bottle</b>			Blood culture bottle, white cap  <b>Minimal volume:</b> 5mL of BAL, 6mL of CSF, 1-5 mL of bone marrow or blood

**TO AVOID SAMPLE REJECTION - PLEASE FOLLOW THE INSTRUCTIONS BELOW:**

**\* SEND THE SAMPLE IN CORRECT CONTAINER**

**\* Sample MUST be labelled with 4 identifiers (District number or NHS number, Surname, Forename, DOB)**

**\* Request form identifiers MUST match the identifiers on the sample**

**\* Each sample must be placed in a separate, sealed plastic bag, samples that require testing in multiple departments MUST be separated and transported in separate bags**

**\* One test should be requested per request form and one sample sent**

**\* Screw cap MUST be securely sealed to prevent sample's leakage**

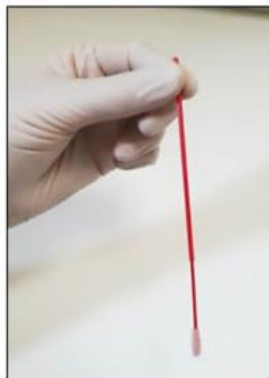
## APPENDIX 1:

# Double eSwab: MRSA screening for nose and groin/perineum only



1. Open the peel pouch and hold with swabs and tube accessible.

Alternatively, the tube can be placed on a flat surface.



2. Take out the pink swab holding **only** the top half of the shaft.



4. Unscrew tube cap, insert swab into the liquid and 'swirl' for 5 seconds.

3. Collect the first sample (groin/perineum).

5. **Discard** the pink swab as tiger waste. Re-cap tube if required.



6. Take out the white swab holding **only** the top half of the shaft.

7. Collect the second sample (nose).



8. Unscrew tube cap, insert the swab into the tube and snap off at marked break point.

9. Discard the remaining plastic shaft.



10. Re-cap the tube with the white swab end and liquid inside.

**Note:** Swab can be dampened with one drop of sterile saline before use if required.

Do not use the liquid from the e-swab as the whole amount is needed for the test.

## APPENDIX 2:

