

**Payment Status:**

NHS

Private

**Tel:**

**Email:**

**Department:**

**Copy report to** (if applicable)**:**

**Hospital** (in full)**:**

**Consultant** (in full)**:**

**Sex:**

**NHS No:**

**DoB:**

**Address/Postcode:**

**Forename:**

**Surname:**

**Hospital No:**

**Referring Clinician**

**Patient Details**

**CNS Tumour Test Request Form**

**North West Genomic Laboratory Hub (MANCHESTER), Manchester Centre for Genomic Medicine (MCGM)**

**CLINICAL DETAILS PLEASE INCLUDE A COPY OF THE PATHOLOGY REPORT**

Pathology Laboratory Hospital/Trust: Pathology block/sample no.:

Sampling Date:

Genomic Request date:

**PLEASE COMPLETE AND FORWARD TO THE PATHOLOGY LABORATORY HOLDING THE SAMPLE.**

|  |  |  |
| --- | --- | --- |
| Test Name | Test Directory Code | Please tick |
| 1p19q FISH |  |  |
| MGMT promoter hypermethylation |  |  |
| KIAA1549:BRAF fusion |  |  |
| ZFTA:RELA fusion (Previously known as C11orf95:RELA fusion) |  |  |
| EGFRvIII transcript |  |  |
| BRAF codon 600 mutation testing |  |  |
| NGS CNS tumour sub-panel – please circle any genes where analysis is a priority (AKT1; ALK; AR; ATRX; BCOR: BRAF; CDKN2A; CTNNB1; DDR2; EGFR; ERBB2; FGFR3; GNA11; GNAQ; H3F3A; H3F3B; HIST1H3B; HIST1H3C; IDH1; IDH2; KIT; KRAS; LZTR1; MAP2K1; MET; NF2; NRAS; PDGFRA; PIK3CA; PTEN; RET; SMARCB1; SMARCE1; SMARCA4; STK11; TERT (including promoter); TP53; VHLCNVs: CDKN2A, CDKN2B, ERBB2, EGFR, MET, PTEN, TP53 |  |  |
| Methylation arrays (please send an additional 4 x 5uM unmounted sections) – NB currently on research basis only |  |  |
| Fusions – NGS fusion panel inc NTRK and FGFR  |  |  |

\*For full details of genes covered see national genomic cancer test directory (<https://www.england.nhs.uk/publication/national-genomic-test-directories/>).

**PATHOLOGY LABORATORY – please complete**

***Please note 2 tubes of curls are required for all testing.*** *For sample requirements please see reverse or* [*https://mft.nhs.uk/nwglh/*](https://mft.nhs.uk/nwglh/)

**Please circle or state the approximate neoplastic cells (%) in the sample sent for analysis** *(this information is important in reducing the risk of false negative results).*

 **Overall neoplastic cell content % #***Neoplastic cells in marked area \_\_\_\_\_\_\_\_%*

#Where overall neoplastic cell content <20% and macrodissection would enhance % of neoplastic cells, please send slide mounted sections with corresponding marked H&E stained slide.

|  |  |  |
| --- | --- | --- |
| 1-5# | 6-10# | 11-20# |
| 20-50 | 50-75 | >75 |

**INFORMATION FOR PATHOLOGY LAB (ALL SAMPLES)**

* Formalin fixed paraffin embedded (FFPE) material should be reviewed by a histo/cyto-pathologist to identify areas containing neoplastic cells and determine suitability for testing.
* Sections should be cut under conditions that prevent cross contamination from other specimens.
* Scrolls should be sent in a sterile tube labelled with **at least 2 patient identifiers, one of which should be the pathology sample number.** Containers and slides should also be labelled with **at least 2 patient identifiers one of which should be the pathology sample number.**
* For each additional test indicated to need additional material please send an additional tube of scrolls.
* Please avoid baking slides or heating samples
* Please send appropriate corresponding paperwork with the samples
* Please contact the laboratory for additional guidance or if you are unsure whether a sample is suitable
* Curls should be sent in 2 ml screw-cap micro tubes where possible. Please contact the lab for more details

**FISH TESTS**

* Prepare 4 unstained sections (4µm thick) floated on the surface of a purified water bath set at 40oC (+/-2oC).
* Mount on positively charged slides and allow to air-dry
* Also include 1 H&E slide with regions enriched for neoplastic cells marked by a Pathologist along with an estimate of neoplastic cell content in the marked area(s)

Complete Sections 1-3 of request form (available for download from https://mft.nhs.uk/nwglh/)

Oncologist/MDT

Pathology Laboratory

FFPE Block

≥20% neoplastic cell content (NCC) overall

2 tubes of 5x10um sections as curls

Yes

No

<20% NCC overall but ≥20% NCC in marked area

10x10um sections mounted on slides (no coverslips) with accompanying H&E slide marked with area for macrodissection

Send to: North West Genomic Laboratory Hub – Manchester Site, Manchester Centre for Genomic Medicine Sample Reception (6th Floor),

St Mary’s Hospital,

Oxford Road,

Manchester, M13 9WL

No

Yes

Sample may not be suitable for testing – please contact the laboratory

Please note our laboratory policy is to discard surplus FFPE material which is not required for genomic testing. This includes H&E stained slides. In the exceptional case where the diagnostic H&E stained slide has been sent and requires return, please email the laboratory (mft.genomics@nhs.net) at the earliest opportunity and within 6 months of sending to request its return.